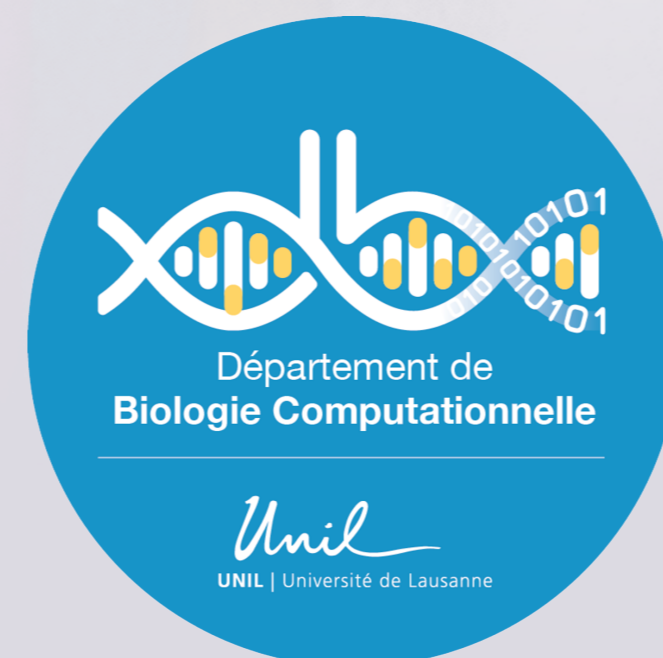


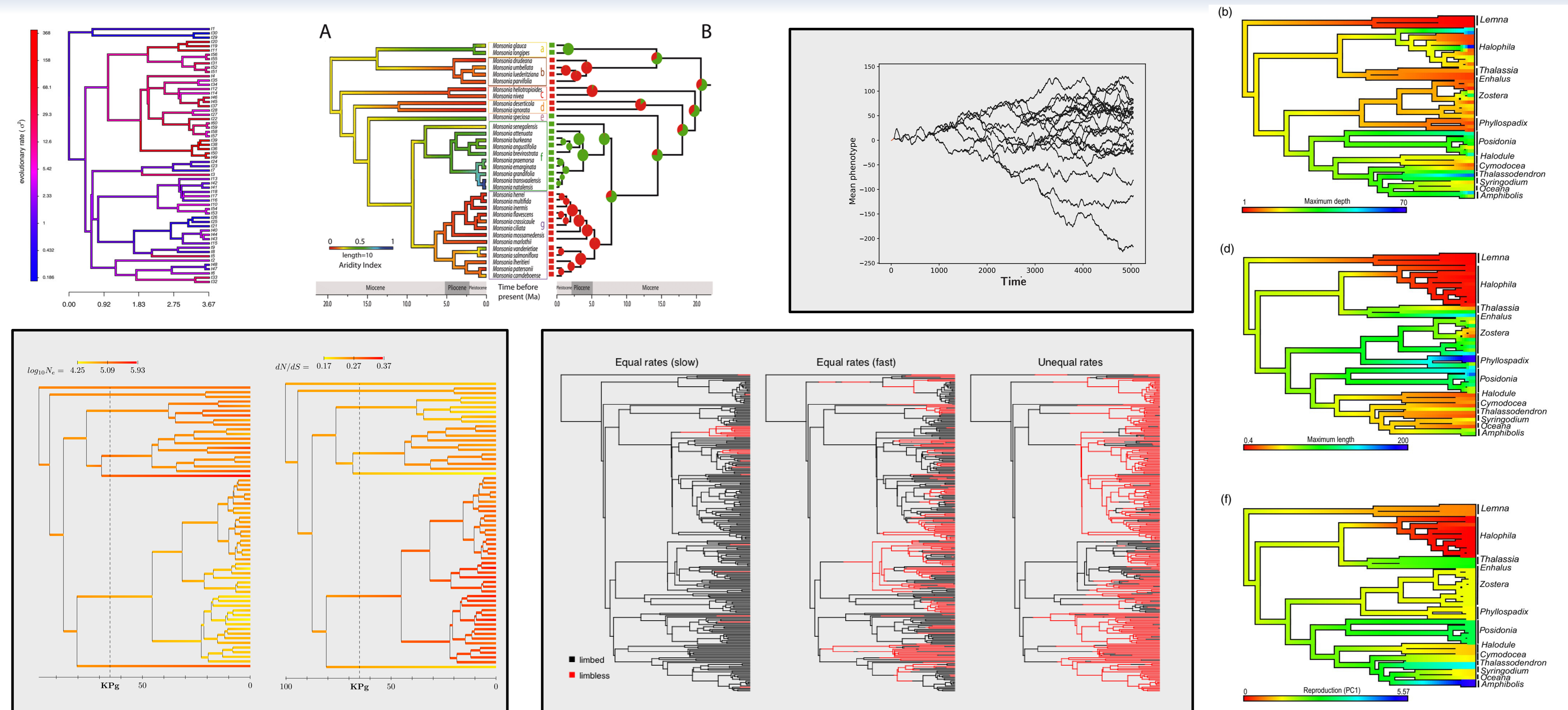
Measuring trait variation in unit of nucleotide changes to disentangle neutral and adaptive processes

Thibault Latrille, Théo Gaboriau, Nicolas Salamin
Université de Lausanne



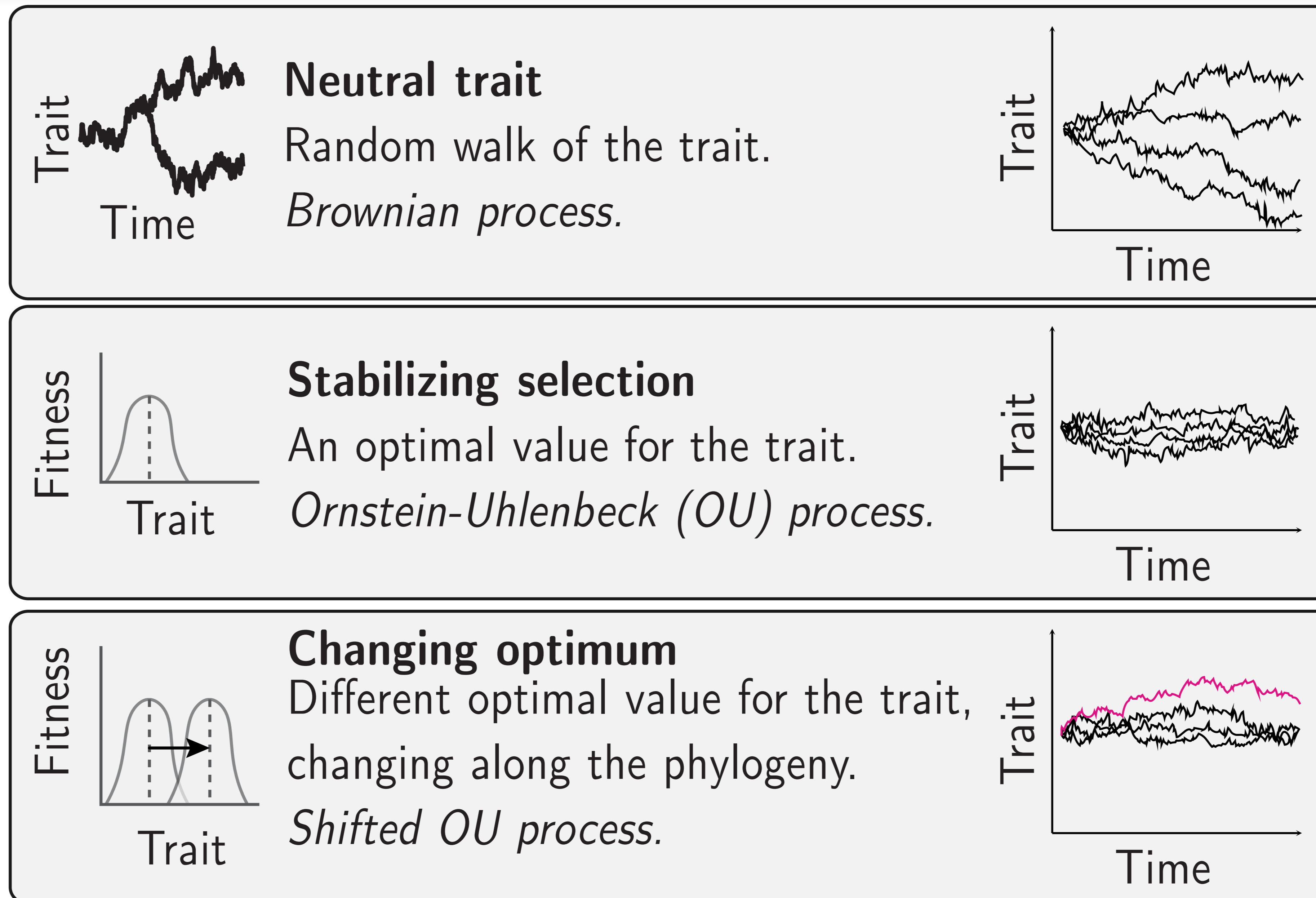
In which unit is measured trait variation?

At the phylogenetic scale, in unit of time.



How to disentangle neutral and adaptive processes?

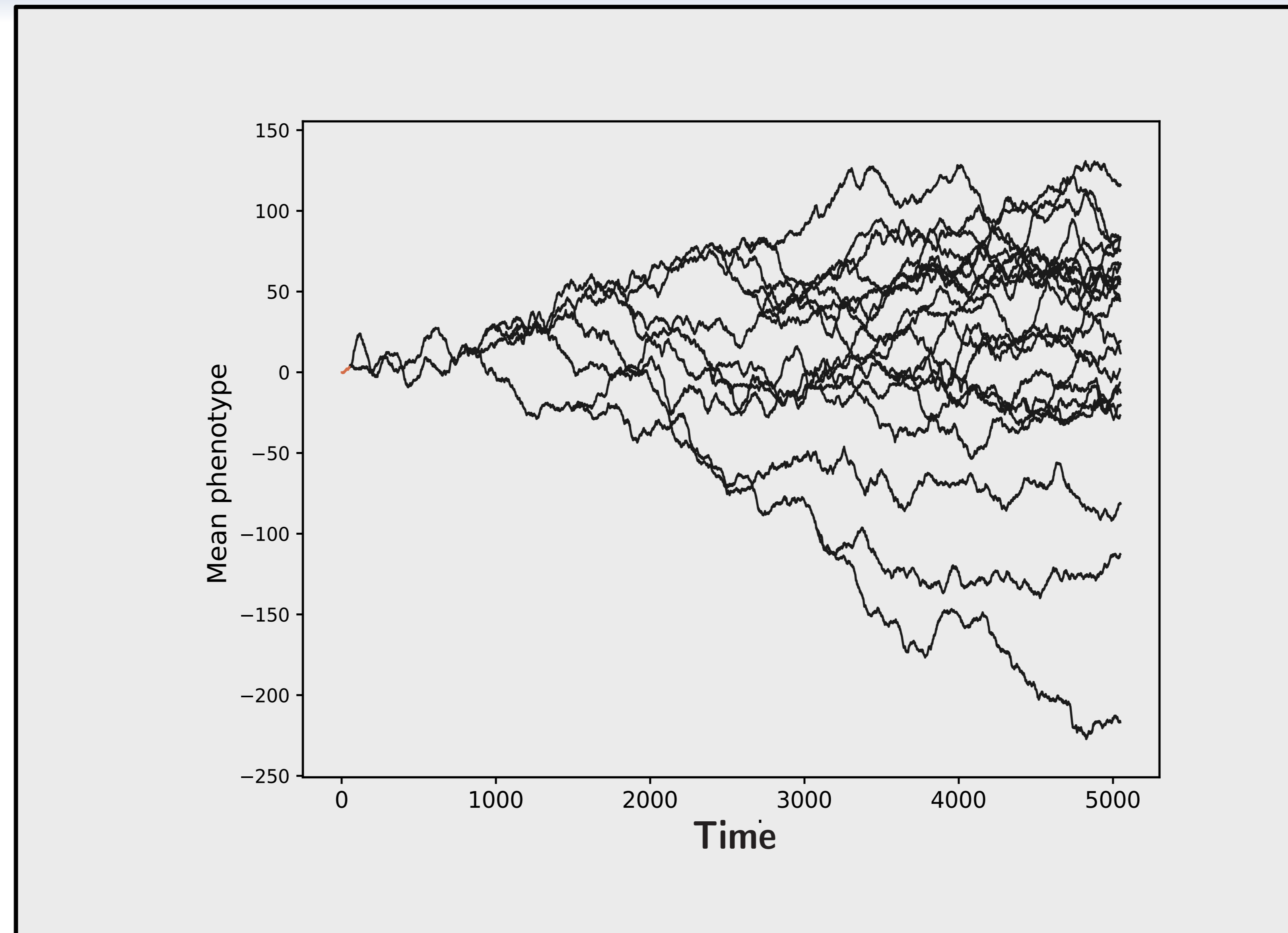
By fitting different models describing different evolutionary scenarios.



Typically, the **best fit of a Brownian process** would conclude that the trait changes are due to **neutral evolution** or “genetic drift”.

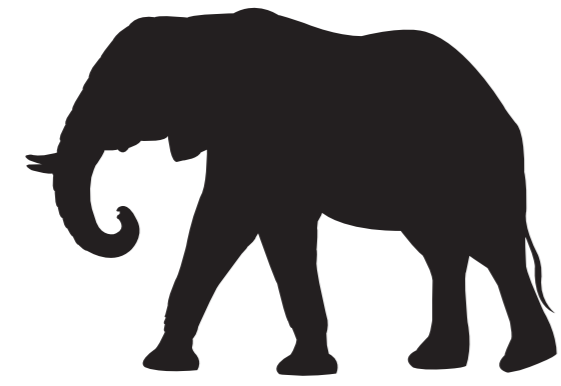
But we have a problem.

The best fit of a Brownian process is **NOT** a proof of neutral evolution.

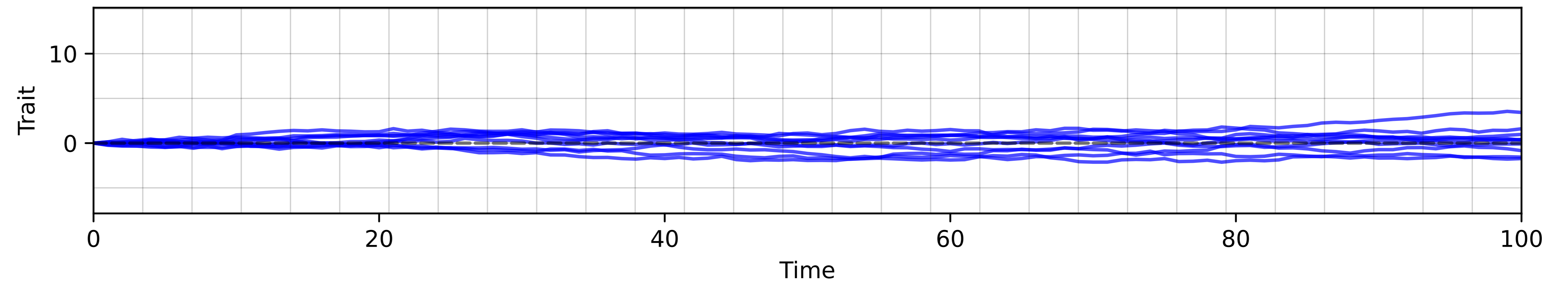


- A neutral trait will follow a Brownian process*. (*under some conditions).
- But a trait under a **constantly changing optimum** might follow a **Brownian process** too.

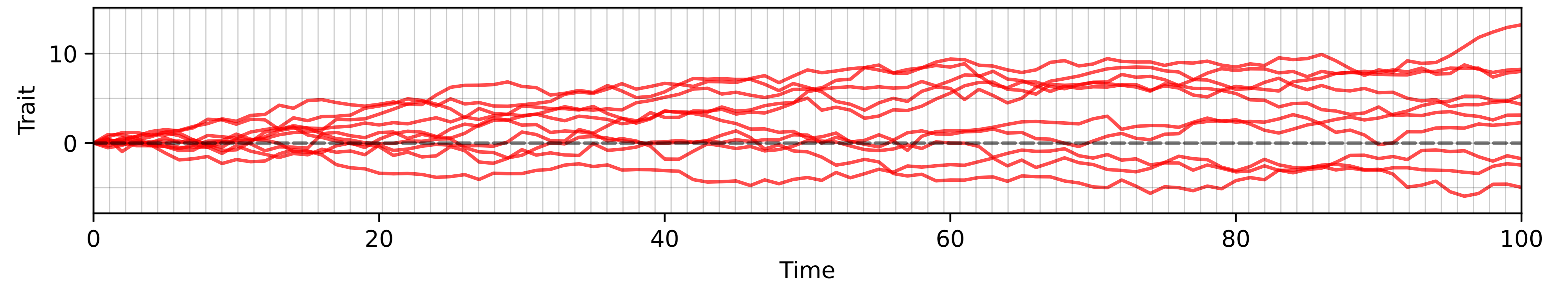
Is time the unit on which is measured the rate of a Brownian.
No, it is the number of generations (for a neutral trait).



Long generation time.
Fewer generations.



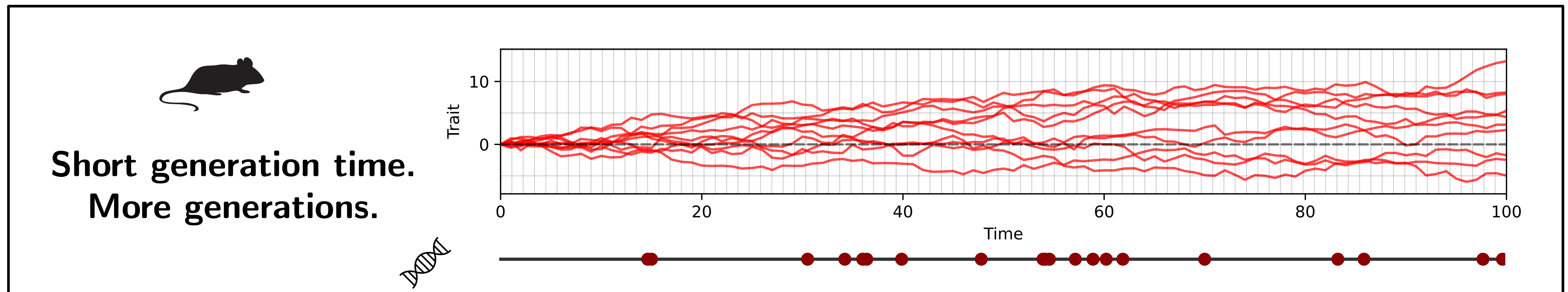
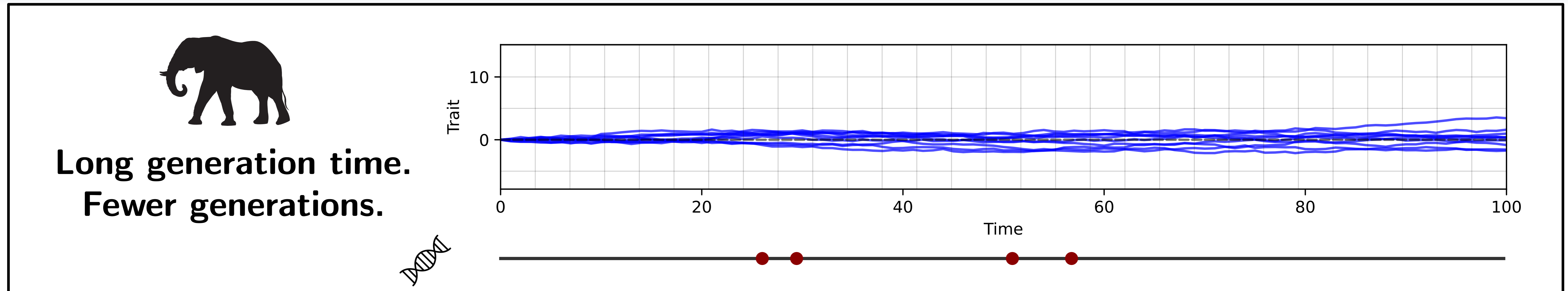
Short generation time.
More generations.



- A **neutral trait** will follow a Brownian process in unit of **generations**.

Is time the unit on which is measured the rate of a Brownian.

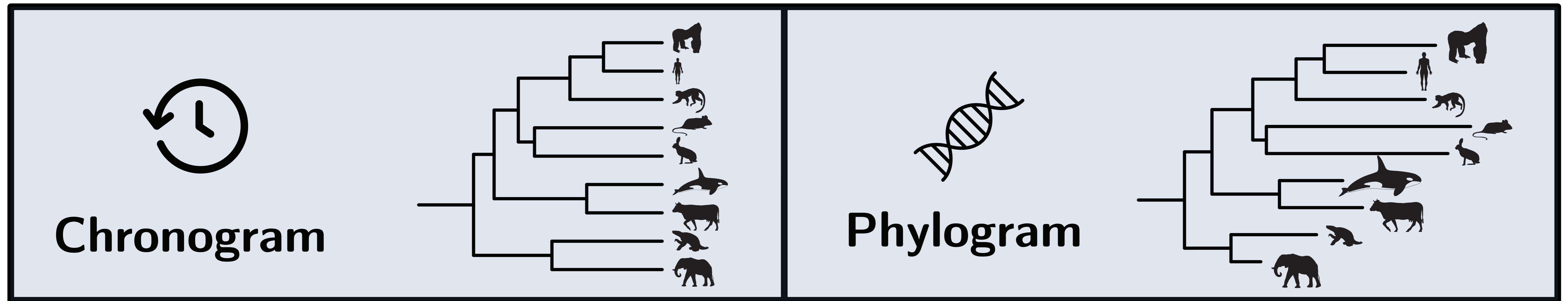
No, it is the number of generations (for a neutral trait).



- A **neutral trait** will follow a Brownian process in unit of **generations**.
- The **number of generations** is proportionnal to **sequence divergence** (.

Can branch lengths be proportionnal to the number of generations?

By using phylogram (number of substitutions) instead chronogram (time).



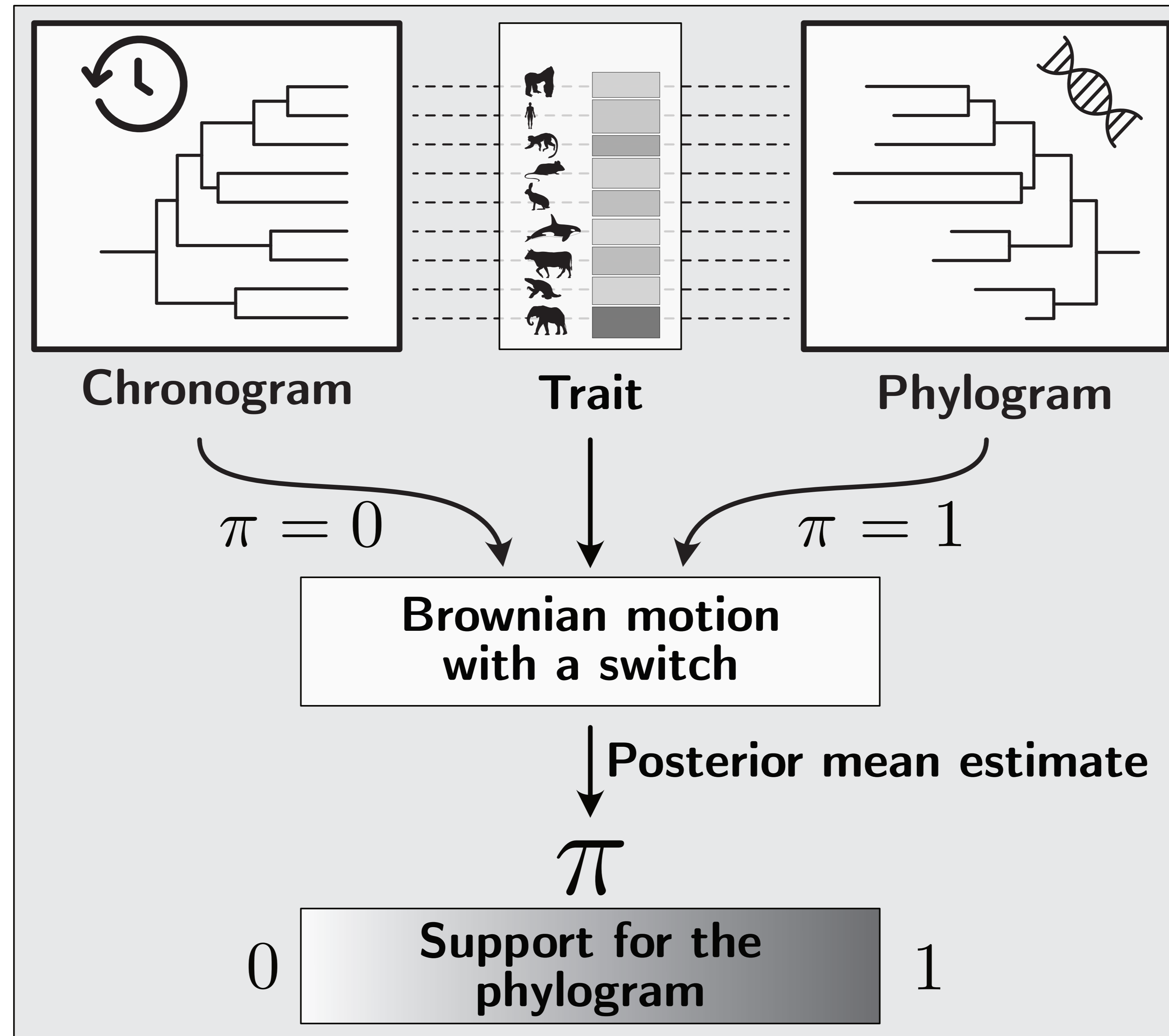
🕒: Chronogram in unit of time.

🧬: Phylogram in unit of expected substitutions per site.

A **neutral trait** will fit a Brownian process better a on a **phylogram** than on a chronogram.

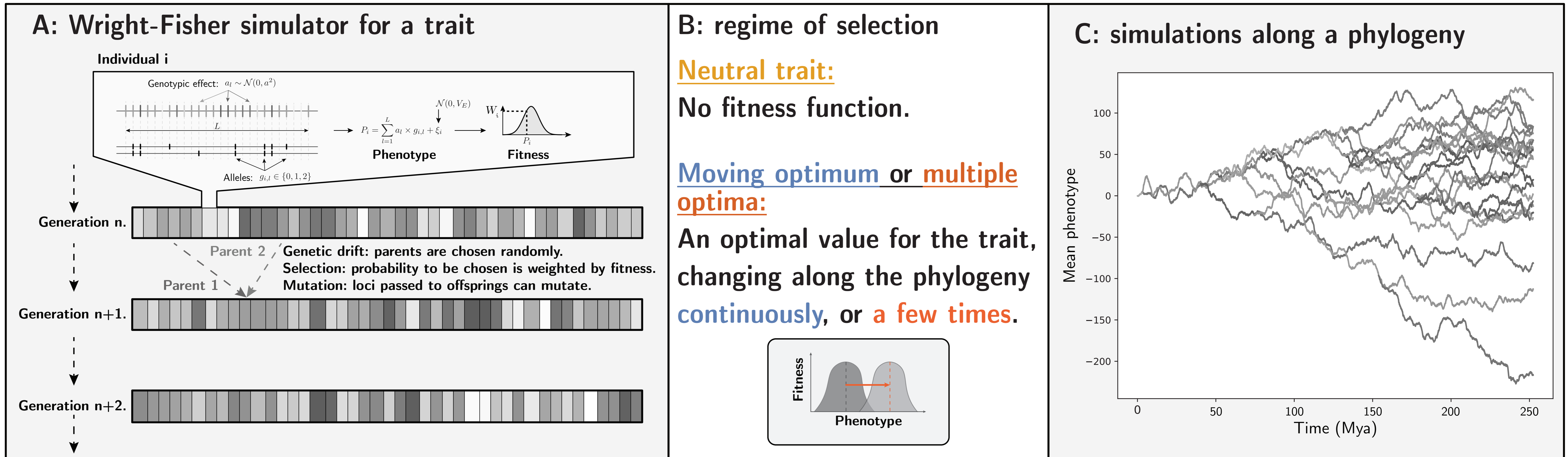
Is a Brownian process best supported on a phylogram?

We can derive a model estimating the support for a phylogram.



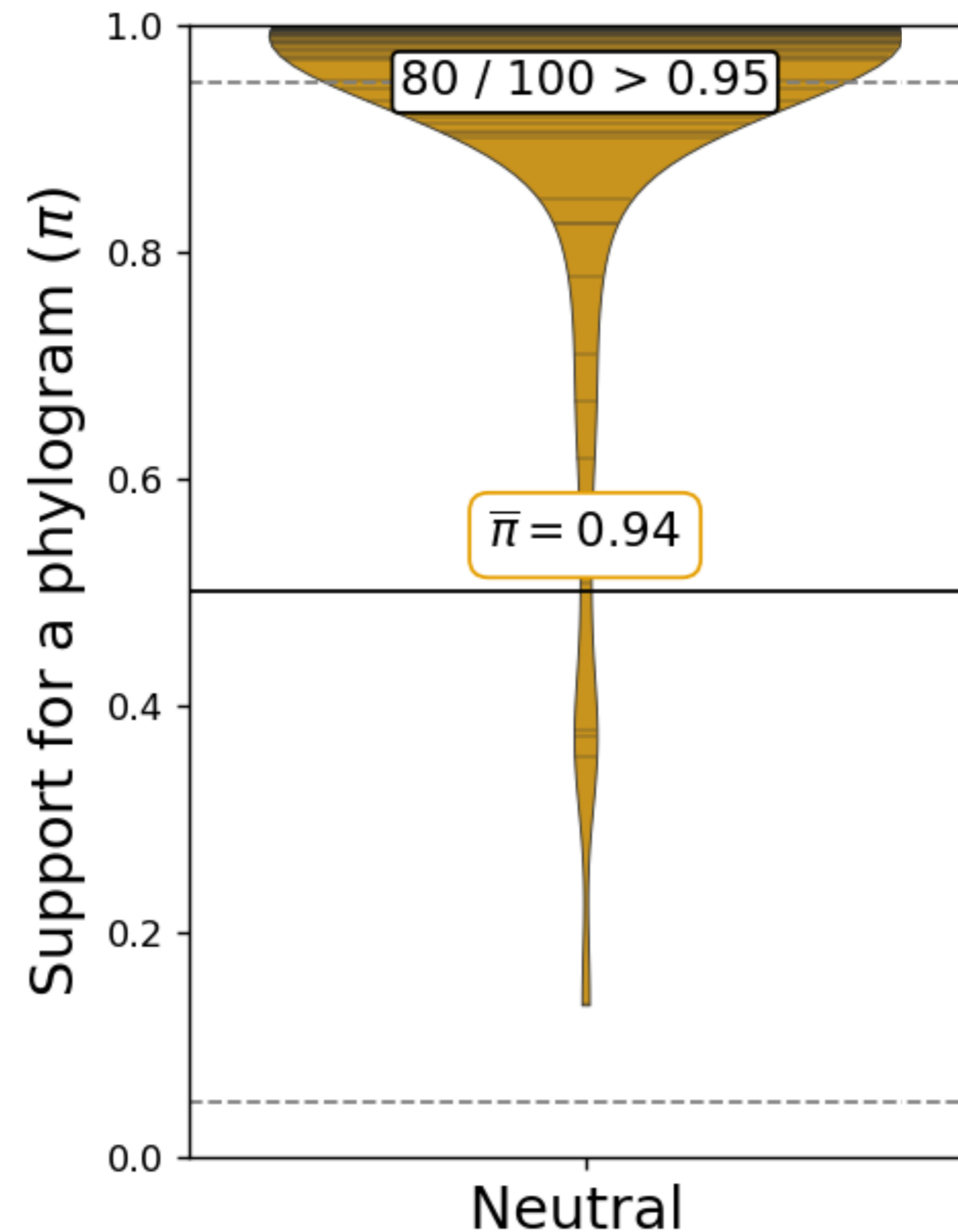
Can we test this model estimating the support for a phylogram?

Using simulations for the different evolutionary scenarios.



Can the support for a phylogram be estimated?

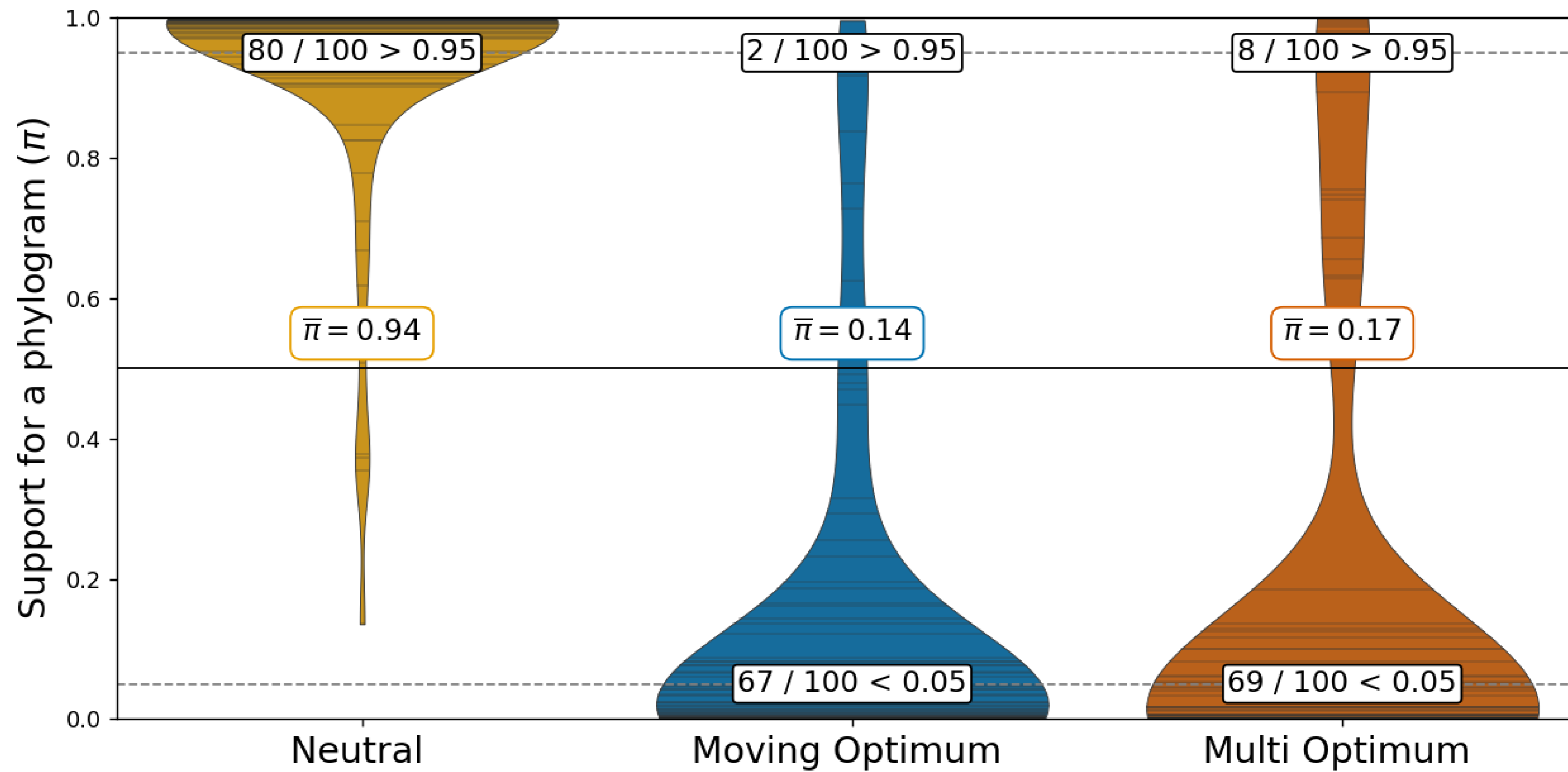
Yes, and it is high for trait simulated under neutral evolution.



- **Support for phylogram** is high if the trait is **neutrally evolving**.

Can the support for a chronogram be estimated?

Yes, and it is high for trait under selection.

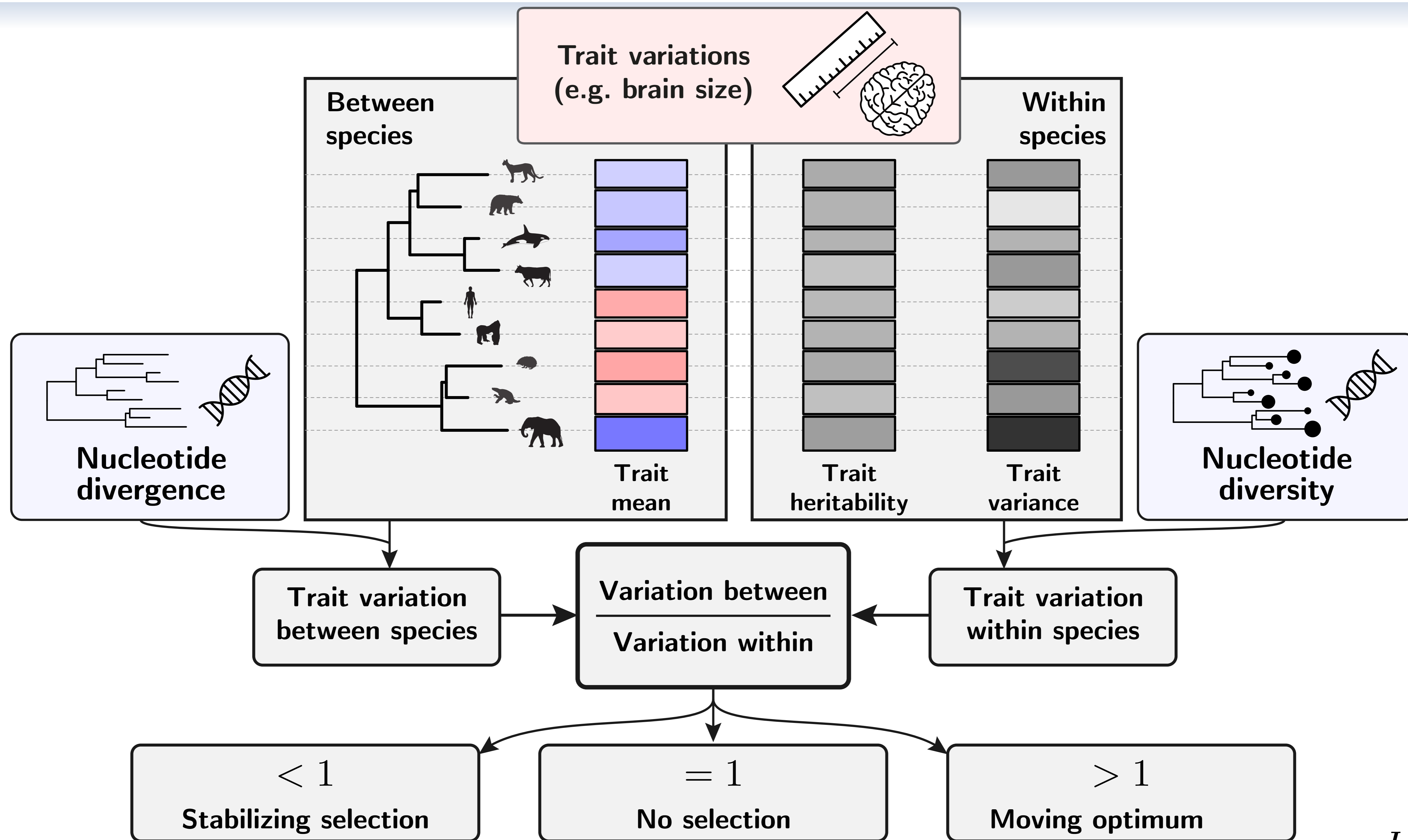


- Support for phylogram is high if the trait is **neutrally evolving**.
- Support for chronogram is high if the trait is **tracking a changing optimum**.

Measuring morphological variation in unit of nucleotide changes to disentangle neutral and adaptive processes

- Changes of a trait **tracking a changing optimum** is better supported on a chronogram (🕒).
- Changes of a **neutral trait** is better supported on a phylogram (🧬).
- If one **claims that a trait is neutrally evolving**, this should be assessed by comparing the **support for a phylogram (🧬) over a chronogram (🕒)**.
- Can we do better and use both between and within species variation?

Can we derive a neutrality index using between and within species variation? By normalising trait variation using nucleotide changes.



Latrille *et al.*
J. Evol. Biol., 2024

How to derive formally a neutrality index?

Similar to dN/dS ratio, or Qst-Fst, but for a trait at the phylogenetic scale.

Within species variation:

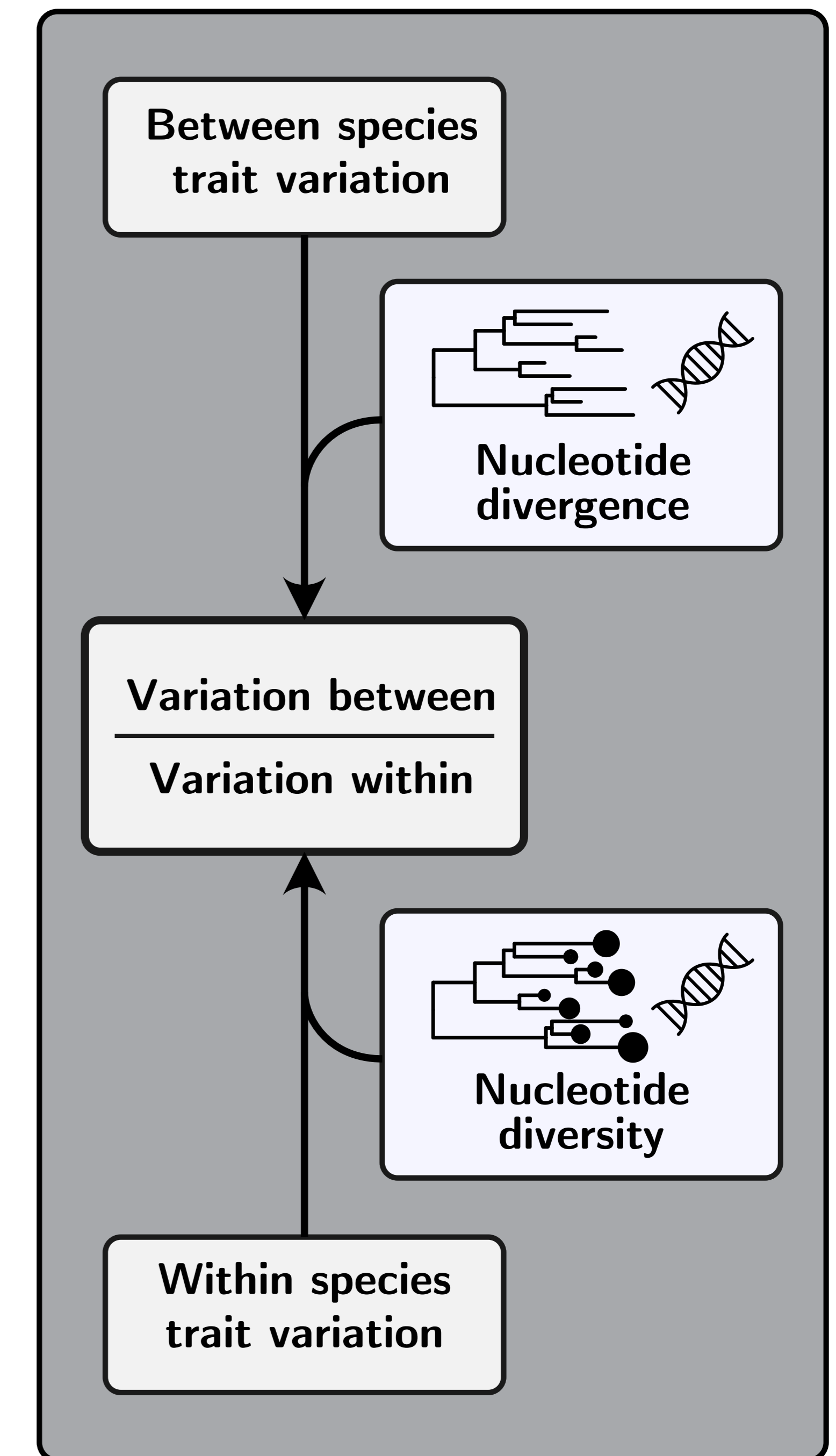
- V_P : phenotypic variance.
- h^2 : heritability for the trait.
- π : mutations per site between two haplotypes.

Between species variation:

- $\text{cov}(\bar{P}_i, \bar{P}_j)$: covariance in mean trait value between a pair of species.
- d : shared nucleotide divergence between a pair of species.

$$\begin{cases} \sigma_W^2 \stackrel{\text{def}}{=} \frac{V_P \cdot h^2}{\pi} = \frac{4N_e \cdot \mu \cdot \sigma_M^2}{4N_e \cdot \mu} = \sigma_M^2, \\ \sigma_B^2 \stackrel{\text{def}}{=} \frac{\text{cov}(\bar{P}_i, \bar{P}_j)}{4d} = \frac{4t \cdot \mu \cdot \sigma_M^2}{4t \cdot q} = \sigma_M^2. \end{cases} \implies \sigma_B^2 / \sigma_W^2 = 1.$$

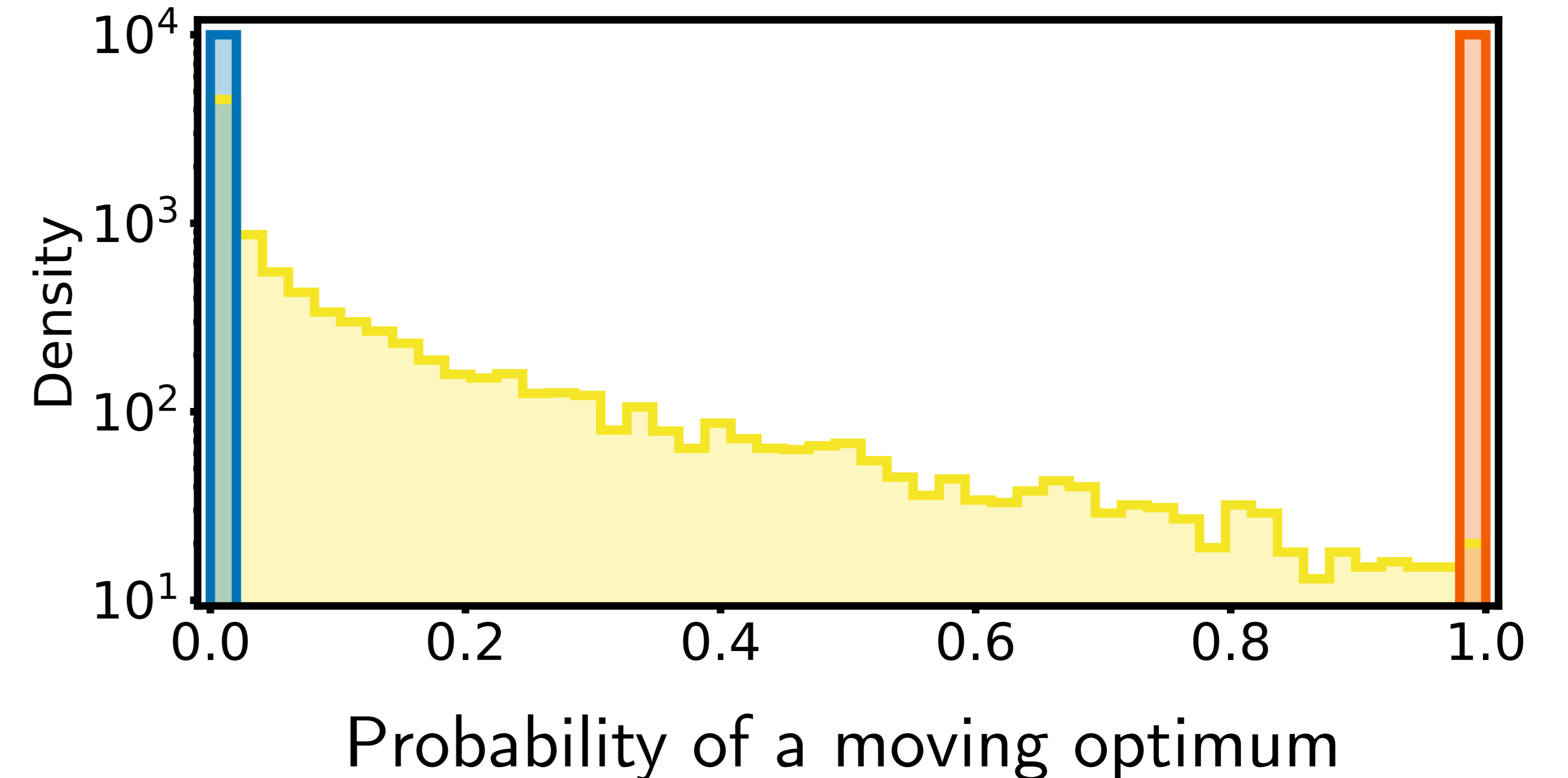
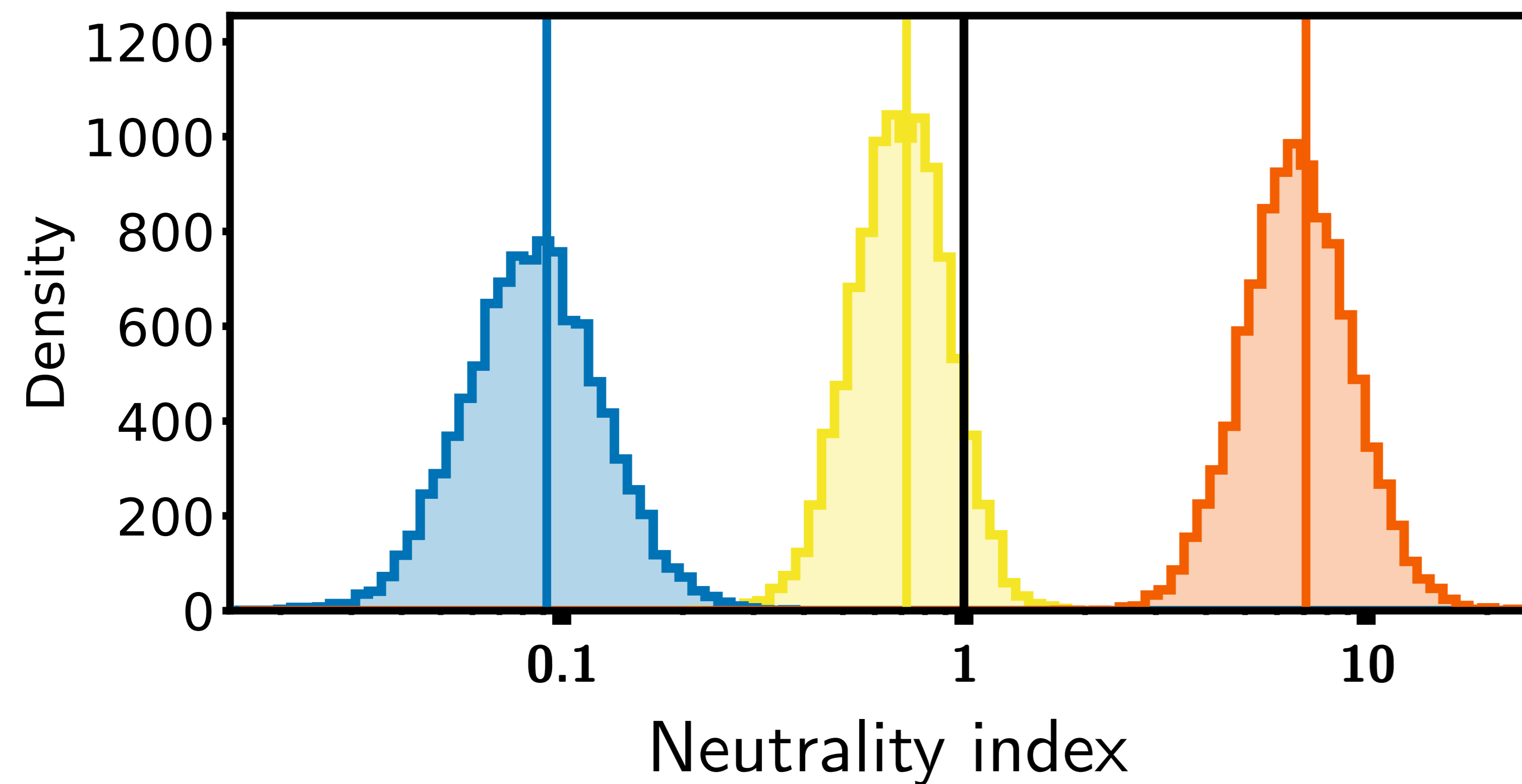
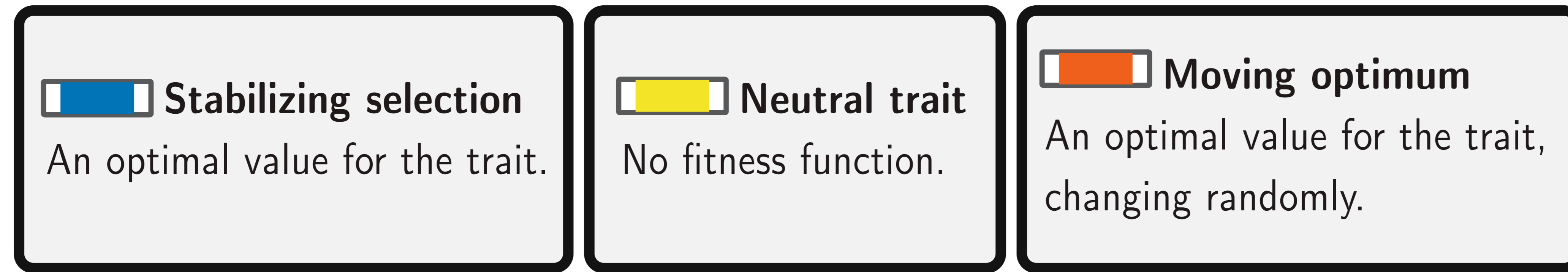
- μ : mutation rate per generation.
- q : substitution rate per generation.
- N_e : effective population size.
- σ_M^2 : effect on the trait per mutation.



Latrille *et al.* (*J. Evol. Biol.*, 2024)

Can we test such neutrality index?

Using simulation under different evolutionary scenarios.



- A neutrality index disentangling neutral and adaptive processes acting on trait.

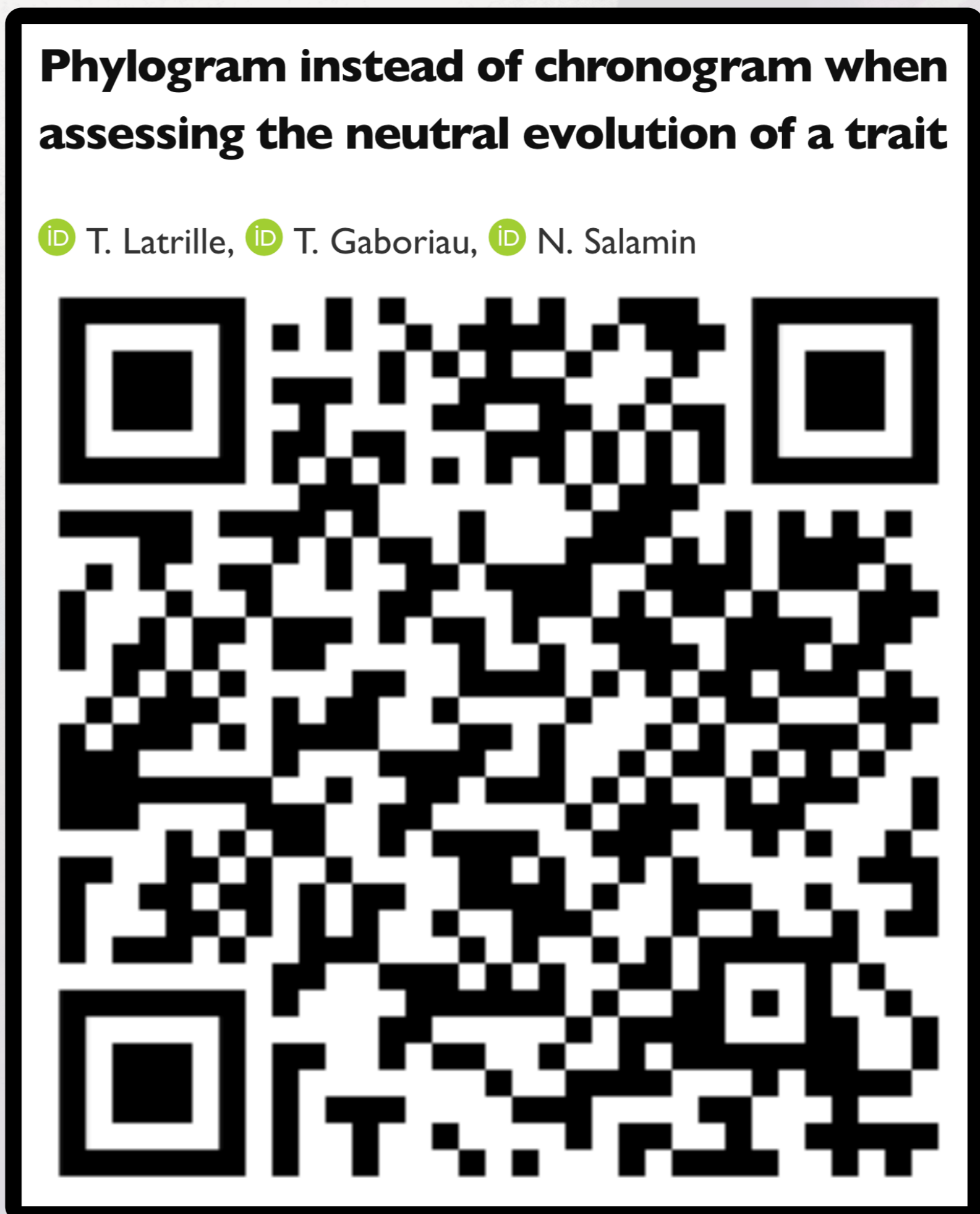
Latrille *et al.* (*J. Evol. Biol.*, 2024)

Nucleotide changes allow disentangling neutral and adaptive processes by contrasting trait variation:

- in unit of **nucleotide divergence** instead of **time**.
- at the between species level (**divergence**) and within species (**diversity**).

Théo Gaboriau
Nicolas Salamin

*Computational
Phylogenetics
Group*

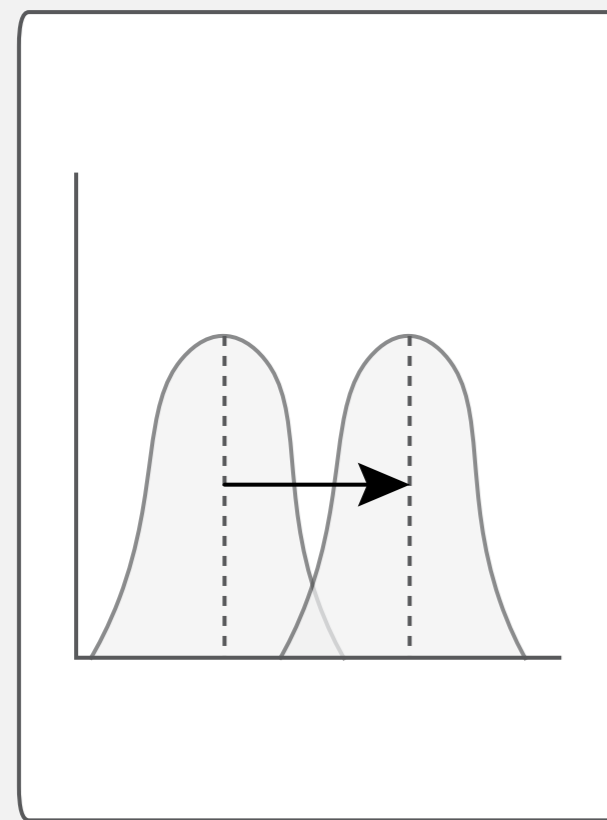


What is the expected rate of evolution for a neutral trait?

Can we derive a test of selection

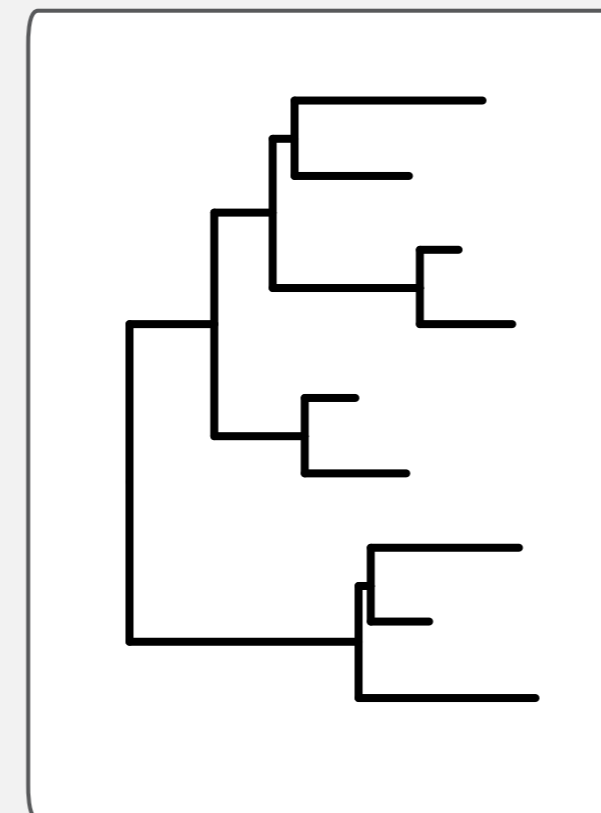
Quantitative-genetics across populations.

→ How to adapt Q_{ST} - F_{ST}
methods across species?



Phylogenetic DNA evolution.

→ How to derive a d_N/d_S
ratio but for a trait instead of
protein coding DNA sequences?



Contrast polymorphism & divergence.

→ How to adapt McDonald &
Kreitman test ($d_N/d_S > p_N/p_S$) for
trait changes along a phylogeny?

